Evidence for Hydrogen Bonding Effects in the Iron Ligand Vibrations of Carbonmonoxy Myoglobin

Masashi Unno,^{†⊥} James F. Christian,[†] John S. Olson,[‡] J. Timothy Sage,[†] and Paul M. Champion^{*,†}

Department of Physics, Northeastern University Boston, Massachusetts 02115 Department of Biochemistry and Cell Biology Rice University, Houston, Texas 77005-1892

Received September 18, 1997 Revised Manuscript Received December 30, 1997

The Fe-CO unit in heme proteins has been used as a sensitive structural probe of the heme active site. Measurement of C-O stretching (ν_{C-O}), Fe-CO stretching (ν_{Fe-CO}), and Fe-C-O bending (δ_{FeCO}) frequencies by vibrational spectroscopies has provided detailed information about protein-ligand interactions.¹ Recent resonance Raman (RR) and infrared (IR) studies of mutant myoglobin (Mb) and model heme compounds have demonstrated that the $v_{\text{Fe}-\text{CO}}$ and $v_{\text{C}-\text{O}}$ frequencies are mainly determined by electrostatic interactions in the distal pocket,^{1,2a} while variations in FeCO geometry are not significant.^{2b} The open conformation of MbCO (A_0 conformer), where the distal histidine (His64) is displaced out of the heme pocket and the bound CO is in a less polar environment, shows the $v_{\text{Fe}-\text{CO}}$ and $v_{\text{C}-\text{O}}$ at 490 and 1965 cm^{-1} , respectively.^{3,4} The closed conformation (A₁ and A₃ conformers), where His64 is present in the heme pocket adjacent to the bound ligand and the CO is in a more polar environment, shows the $\nu_{\text{Fe}-\text{CO}}$ and $\nu_{\text{C}-\text{O}}$ at 510–520 and 1945–1935 cm⁻¹, respectively. Replacement of His64 by aliphatic or aromatic amino acids produces $v_{\text{Fe-CO}}$ and $v_{\text{C-O}}$ bands whose frequencies are similar to the corresponding values when the distal histidine is in the open conformation in native MbCO.1 In this communication we report RR spectroscopic evidence that suggests a hydrogen bond in the A₁ conformation of MbCO between the iron-bound CO and the distal histidine. This implies that a proton exists at the N_{ϵ} position in the A₁ conformer of MbCO and contributes to the increased $v_{\text{Fe-CO}}$ and decreased $v_{\text{C-O}}$ compared to the A₀ conformation.

We examined⁵ the D_2O effects on the resonance Raman spectra of native and five distal histidine mutants (His64 \rightarrow Ala, Val, Leu, Ile, and Phe) of sperm whale MbCO. Figure 1A shows the low-frequency resonance Raman spectra of native MbCO in

* To whom correspondence should be addressed.

 (1) (a) Ray, G. B.; Li, X.-Y.; Ibers, J. A.; Sessler, J. L.; Spiro, T. G. J. Am. Chem. Soc. **1994**, 116, 162–176. (b) Li, T.; Quillin, M. L.; Phillips, G. N., Jr.; Olson, J. S. Biochemistry **1994**, 33, 1433–1446. (c) Ling, J.; Li, T.; Olson, J. S.; Bocian, D. F. Biochim. Biophys. Acta **1994**, 1188, 417–421. (d) Anderton, C. L.; Hester, R. E.; Moore, J. N. Biochim. Biophys. Acta **1997**, 1338, 107–120. (e) Decatur, S. M.; Boxer, S. G. Biochem. Biophys. Res. Commun. **1995**, 212, 159–164.

(2) (a) Oldfield, E.; Guo, K.; Augspurger, J. D.; Dykstra, C. E. J. Am. Chem. Soc. **1991**, 113, 7537–7541. (b) Ivanov, D.; Sage, J. T.; Keim, M.; Powell, J.; Asher, S.; Champion, P. M. J. Am. Chem. Soc. **1994**, 116, 4139–4140.

(3) Yang, F.; Phillips, G. N., Jr. J. Mol. Biol. 1996, 256, 762-774.

(4) (a) Morikis, D.; Champion, P. M.; Springer, B. A.; Sligar, S. G.
 Biochemistry 1989, 28, 4791–4800. (b) Zhu, L.; Sage, J. T.; Rigos, A. A.;
 Morikis, D.; Champion, P. M. J. Mol. Biol. 1992, 224, 207–215. (c) Ramsden,
 J.; Spiro, T. G. Biochemistry 1989, 28, 3125–3128.

(5) Resonance Raman spectra were obtained on a Spex 1870B spectrometer equipped with a liquid nitrogen cooled CCD detector (Princeton Instruments, Inc.) and a notch filter (Kaiser Optical Systems, Inc.). Samples were excited with the 413.1 nm line available from a Coherent Innova 300 krypton ion laser. All spectra were taken at room temperature, and the laser power was about 1 mW. The measurements were made on samples contained in a quartz

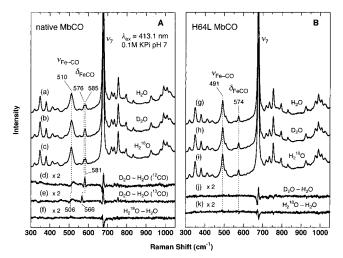


Figure 1. Low-frequency region of resonance Raman and difference spectra of native (A) and the His64 \rightarrow Leu mutant (B) of sperm whale myoglobins in 0.1 M potassium phosphate buffer, pH or pD 7.0: (a, g) 100% H₂O; (b, h) 90% D₂O/10% H₂O; (c, i) 90% H₂¹⁸O/10% H₂O; (d, j) D₂O - H₂O for Mb¹²CO; (e) D₂O - H₂O for Mb¹³CO; (f) H₂¹⁸O - H₂O.

buffered H₂O, D₂O, and H₂¹⁸O solutions, as well as D₂O – H₂O and H₂¹⁸O – H₂O difference spectra. The D₂O – H₂O difference spectrum (trace d) shows changes around 510 and 580 cm⁻¹. The shift of ν_{Fe-CO} at 510 cm⁻¹ is approximately +1 cm⁻¹. When we use ¹³CO as a ligand, the derivative pattern in the difference spectrum is downshifted by 4 cm⁻¹ (trace d \rightarrow e), demonstrating that the observed D₂O-induced shift originates from the Fe–CO stretching mode.⁶

The D₂O-induced spectral change in the vicinity of the doublet bands near 580 cm⁻¹ involves both an upshift and an intensity increase of the 576 cm⁻¹ mode, leading to a loss of the doublet structure (trace b) and a prominent difference feature (trace d). Upon ¹³CO substitution, the doublet feature at 580 cm⁻¹ appears as a singlet at 562 cm⁻¹ (suggestive of a Fermi resonance), and the H/D exchange again upshifts the frequency (~2 cm⁻¹) and increases the intensity so that the difference spectrum yields a feature at 566 cm⁻¹ (trace e, see also the figure in the Supporting Information). In contrast to Hirota et al.,⁷ we agree with the previous assignments⁸ for the band at 576 cm⁻¹ as the Fe–C–O bending mode having a doublet feature that arises from a Fermi resonance with a nearby porphyrin mode.

Analogous experimental data for the His64 \rightarrow Leu MbCO (Figure 1B) do not exhibit any detectable H/D isotope substitution effects, nor do the other His64 mutants examined in this study. This demonstrates that His64 contributes to the observed D₂O effect for the native MbCO. The negligible changes due to H₂¹⁸O in the native and mutant samples serve as an additional control that probes for potential effects associated with water in the distal pocket.

Northeastern University.

[‡] Rice University.

 ¹ Present address: Institute for Chemical Reaction Science, Tohoku University, Sendai 980, Japan.
 (1) (a) Ray, G. B.; Li, X.-Y.; Ibers, J. A.; Sessler, J. L.; Spiro, T. G. J.

spinning cell to minimize the contribution of photodissociated MbCO. Native Mb was obtained from Sigma Chemical Co. and used without further purification. The ¹³CO at 99% enrichment (~10% ¹⁸O) was from Cambridge Isotope Laboratories, Inc. Preparation of mutant Mb has been described elsewhere.^{1b}

⁽⁶⁾ Hirota et al.⁷ examined the H/D substitution effects on the $\nu_{\rm Fe-CO}$ and $\delta_{\rm FeCO}$ Raman bands for horse Mb and human Hb. Although a change at the stretching region can be discerned, their signal-to-noise level did not allow them to recognize it as a real shift.

⁽⁷⁾ Hirota, S.; Ogura, T.; Shinzawa-Itoh, K.; Nagai, M.; Kitagawa, T. J. Phys. Chem. **1994**, *98*, 6652–6660.

^{(8) (}a) Tsubaki, M.; Srivastava, R. B.; Yu, N.-T. *Biochemistry* **1982**, *21*, 1132–1140. (b) Hu, S.; Vogel, K. M.; Spiro, T. G. J. Am. Chem. Soc. **1994**, *116*, 11187–11188.

Prior infrared studies have searched for deuterium isotope effects on ν_{C-O} . For example, no effect of H/D exchange on the $v_{\rm C-O}$ mode in Hb was observed in an early experiment.⁹ The CO adducts of horseradish peroxidase¹⁰ and cytochrome cperoxidase¹¹ show a 2.0–2.5 cm⁻¹ downshift in v_{C-O} when the sample is prepared in D₂O. This effect was attributed to either a hydrogen bonding interaction between the CO and a distal residue or protein conformational changes in D₂O. In the following, we will discuss possible explanations for the D₂O effects on the $v_{\text{Fe-CO}}$ and δ_{FeCO} modes of MbCO.

Deuterium substitution of exchangeable protons might alter protein tertiary structure. The 1.4 cm⁻¹ downshift of the Fe-His stretching mode in deoxy Mb upon deuterium substitution is significantly larger than the value expected on the basis of a simple mass effect (0.7 cm⁻¹),¹² suggesting that deuteration of labile protons could alter the Fe-His bond.¹³ While such a change on the proximal side could affect the vibrational character of the FeCO unit, it fails to explain the lack of H/D exchange effects on mutants of MbCO whose His64 is replaced with aliphatic or aromatic residues (Figure 1B). We therefore suggest that a weak hydrogen bond between His64 and the iron-bound CO makes the Fe-CO stretching and Fe-C-O bending modes sensitive to the H/D exchange. This explanation accounts for the lack of effects on the $v_{\text{Fe}-\text{CO}}$ and δ_{FeCO} modes of His64 mutants MbCO. Kinetic studies on mutant Mbs have shown that the replacement of His64 increases the CO dissociation rate by a factor of 2-3, ^{1b,15a,b} while replacement of His64 with aliphatic residues results in an ~1000fold increase in the O₂ dissociation rate.^{15a,16} These results suggest that hydrogen bonding to His64 stabilizes bound CO by only ${\sim}2$ kJ mol⁻¹ whereas bound O₂ is stabilized by ~ 17 kJ mol⁻¹.^{15b,16,17a}

The proposed hydrogen bond between N_{ϵ} of His64 and the carbonyl oxygen conflicts with the neutron crystal structure,¹⁸ where no proton was found on N_{ϵ} . However, recent spectroscopic, mutagenesis, and theoretical studies suggest a positive polar interaction between His64 and bound CO,^{1,17} consistent with a protonated $N_{\varepsilon}^{,1,15,17}~$ Li and Spiro^{19b} have interpreted the inverse correlation between the ν_{C-Q} and ν_{Fe-CQ} frequencies^{1a,b,19} in terms

(12) Argade, P. V.; Sassaroli, M.; Rousseau, D. L.; Inubushi, T.; Ikeda-Saito, M.; Lapidot, A. J. Am. Chem. Soc. 1984, 106, 6593-6596.

(13) A further analysis¹⁴ suggests that an internal mode of the histidine participates in the Fe-His motion and leads to better agreement with the observed D2O-induced shift. Thus, the observed shift of the Fe-His mode is not necessarily due to a tertiary structure change that affects the Fe-His bond strength

strength.
(14) Wells, A. V.; Sage, J. T.; Morikis, D.; Champion, P. M.; Chiu, M. L.;
Sligar, S. G. J. Am. Chem. Soc. 1991, 113, 9655–9660.
(15) (a) Springer, B. A.; Sligar, S. G.; Olson, J. S.; Phillips, G. N., Jr.
Chem. Rev. 1994, 94, 699–714. (b) Olson, J. S.; Phillips, G. N. J. Biol. Inorg.
Chem. 1997, 2, 544–552. (c) Phillips, S. E. V.; Schoenborn, B. P. Nature
1091, 202, 81–82. 1981, 292, 81-82.

(16) Tian, W. D.; Sage, J. T.; Champion, P. M. J. Mol. Biol. 1993, 233, 155-166.

(17) (a) Lai, H. H.; Li, T.; Lyons, D. S.; Phillips, G. N., Jr.; Olson, J. S.; Gibson, Q. H. Proteins: Struct. Function Genet. **1995**, 22, 322–339. (b) Jewsbury, P.; Kitagawa, T. Biophys. J. 1995, 68, 1283–1294. (c) Kushkuley,
 B.; Stavrov, S. S. Biophys. J. 1997, 72, 899–912.

(18) (a) Hanson, J. C.; Schoenborn, B. P. J. Mol. Biol. **1981**, 153, 117–146. (b) Cheng, X.; Schoenborn, B. P. J. Mol. Biol. **1991**, 220, 381–399.

of back-donation from the iron atom. A positive polar interaction, such as hydrogen bonding, increases the back-bonding donation from the Fe d_{π} to the CO π^* orbital, thereby strengthening the Fe-CO bond (i.e., increasing the frequency) and weakening the C-O bond (i.e., decreasing the frequency). In the absence of such interactions (e.g., in His64 mutants or the A₀ conformation of native MbCO), the $v_{\text{Fe-CO}}$ peak appears at 495 cm⁻¹ whereas the peak for ν_{C-O} is found at 1965 cm⁻¹. Thus, the ν_{Fe-CO} and $v_{\rm C-O}$ peaks at 510 and 1945 cm⁻¹ for the A₁ conformation suggest an interaction with a positive electrostatic field, presumably due in part to the protonated N_{ϵ} of His64. This interpretation is supported by infrared crystallographic determination of the C–O orientation in sperm whale MbCO crystals, which places the carbonyl oxygen 2.6–2.9 Å from N_{ϵ} , a distance consistent with hydrogen bonding.20

The effects of H/D exchange on native Mb shown in Figure 1 provide spectroscopic evidence for a hydrogen bond between His64 and iron bound CO. However, the specific mechanism by which D₂O perturbs the Fe-CO stretching and Fe-C-O bending modes remains uncertain. The H/D isotope effect is not a simple mass effect, since a D₂O-induced upshift of $\nu_{\text{Fe}-\text{CO}}$ and δ_{FeCO} was observed. On the other hand, the reduction in the zeropoint energy of the His64 N_{ϵ}-D bond can stabilize the deuterium bond compared to the hydrogen bond. An important mechanism for this stabilization arises from the reduced mean square displacement of the $N_{\varepsilon}\text{--}D$ "wagging" motion.^{21} The resultant strengthening of the CO···D–N_e interaction could "stiffen" the Fe-CO bending and stretching potentials (with concomitant increases in those frequencies). This scenario does not rely on π back-bonding arguments to explain the H/D isotope shifts and is consistent with the weak H/D perturbations of the C-O mode observed in related infrared experiments.²² On a more global scale, small alterations in charge distribution, due to changes in the zero-point width and position of the His64 N_{ϵ} deuterium, could weakly perturb the electric fields and forces felt in the distal pocket, leading to small changes in the orientation of the imidazole side chain and/or the bound ligand that affect the $v_{\text{Fe}=CO}$ and $\delta_{\text{Fe}CO}$ frequencies. The histidine specific nature of the perturbation is suggested by the absence of a H/D exchange effect in the Gln64 MbCO,²³ where the presence of a hydrogen bond is suggested in the oxy form.¹⁵ To examine these possibilities, further RR and IR studies are currently in progress.

Acknowledgment. We are grateful to Anand Kumar for helpful discussions. This work was supported by grants from NIH DK-35090 (P.M.C.), GM 52002 (J.T.S.), GM 35649 (J.S.O.), HL 47020 (J.S.O.), NSF 94-05979 (P.M.C.), the Welch Foundation C-612 (J.S.O.), and the W. M. Keck Foundation (J.S.O.).

Supporting Information Available: Figure showing the Fe-CO stretching and Fe-C-O bending mode region of the resonance Raman spectra of native and His64 \rightarrow Leu mutant carbonmonoxy myoglobin (1 page, print/PDF). See any current masthead page for ordering information and Web access instructions.

JA973293D

⁽⁹⁾ Satterlee, J. D.; Teintze, M.; Richards, J. H. Biochemistry 1978, 17, 1456 - 1462

^{(10) (}a) Smith. M. L.; Ohlsson, P.-I.; Paul, K. G. FEBS Lett. 1983, 163, (10) (a) Sinih, M. E., Sinsson, F.A., Fadi, K. G. Fills Len. 196, 165, 303–305. (b) Uno, T.; Nishimura, Y.; Tsuboi, M.; Makino, R.; Iizuka, T.; Ishimura, Y. J. Biol. Chem. **1987**, 262, 4549–4556.

⁽¹¹⁾ Satterlee, J. D.; Erman, J. E. J. Am. Chem. Soc. 1984, 106, 1139-1140.

^{(19) (}a) Yu, N.-T.; Kerr, E. A. In *Biological Applications of Raman Spectroscopy*; Spiro T. G., Ed.; Wiley-Interscience: New York, 1988; Vol. III, Chapter 2. (b) Li, X.-Y.; Spiro, T. G. J. Am. Chem. Soc. 1988, 110, 6024-6033

⁽²⁰⁾ Sage, J. T.; Jee, W. J. Mol. Biol. 1997, 274, 21-26.

⁽²¹⁾ Scheiner, S.; Cuma, M. J. Am. Chem. Soc. 1996, 118, 1511–1521.
(22) Unno, M.; Sage, J. T. To be submitted for publication.

⁽²³⁾ Unno, M.; Christian, J. F.; Olson, J. S.; Champion, P. M. Unpublished results.